

A Critical Review of the Biotransformation of Octamethylcyclotetrasiloxane (D4) and Decamethylcyclopentasiloxane (D5) in Fish



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Abstract

Biotransformation is an important physiological process whereby a fish can convert a chemical to a more polar form so that it may be eliminated from the whole body. An understanding of the potential for a chemical to be biotransformed provides important information for a bioaccumulation assessment. Octamethylcyclotetrasiloxane (D4) and decamethylcyclopentasiloxane (D5) are widely used in consumer products and industrial applications. These two siloxanes have a high octanol-water partition coefficient ($\log K_{ow} > 6$), which is suggestive of a high aqueous bioconcentration factor (BCF). Several studies employing high performance liquid chromatography demonstrate that D4 and D5 siloxane are biotransformed into more polar metabolites. A third *in vivo* study employed whole body autoradiography (WBA) and found that a bulk of the ^{14}C -D4 and D5 radioactivity was associated with the liver, gall bladder and digestive tract during and after exposure. *In vitro* microsomal studies suggest that ^{14}C -D5 was biotransformed by rainbow trout, while minimal biotransformation was observed with common carp and channel catfish. Using these data-sets, an estimated k_m for D4 and D5 siloxane is $> 0.01 \text{ day}^{-1}$. Based on the available data, there is conclusive evidence that D5 siloxane is biotransformed to more polar metabolites in fish. This biotransformation is important and provides rationale for D4 and D5 biodilution behavior generally observed in aquatic food webs (i.e. a TMF < 1).

State of The Science

D4

- Juvenile rainbow trout ($1.18 \pm 0.16 \text{ g}$ wet weight) were fed ^{14}C -D4 siloxane for 35 d and then allowed to depurate for 42d (Woodburn et al. 2013; Drottar 2007). The dietary route of exposure represents the most likely exposure route for fish to D4. Whole body measurements from the last three sampling points of the uptake period revealed that the fish achieved steady-state as the residue concentrations did not significantly differ over time. An empirical BMF value for D4 siloxane was 0.28. More importantly, whole body autoradiography (WBA) was conducted on Days 1 and 10 of the uptake period and Days 2, 14 and 42 of depuration (Drottar 2007). During uptake, the concentration of ^{14}C radioactivity was highest in the gastrointestinal tract and adipose tissue, while the concentration of ^{14}C radioactivity was highest in the gall bladder, liver and digestive tract during the depuration period. These data are suggestive of ^{14}C -D4 metabolism and elimination.
- Rainbow Trout (500-1000g) were fed ^{14}C - D4 siloxane for five days, which had been mixed into fish feed (Durham et al. 2009). Individual trout were homogenized, extracted with tetrahydrofuran (THF), and then fractionated and analyzed by high performance liquid chromatography (HPLC) to profile potential metabolites. Based on the HPLC metabolite profiling, $\geq 5\%$ of the radioactivity in the fish homogenates were D4 siloxane metabolites.
- Rainbow Trout (967 – 1377g) were administered a single orally gavage dose of 15 mg/kg ^{14}C -D4 siloxane (Springer 2007; Domoradzki et al. 2008). Over 96 hr, trout plasma and urine were sampled to determine the amount of parent D4 siloxane and metabolites in each matrix. At the 96 hr terminal sacrifice, trout bile, digestive tract, liver, and fat were also sampled for parent D4 siloxane and metabolites. HPLC metabolite profiling demonstrated that biotransformation products were present in the plasma, urine, bile and liver (Appendix 2). Parent D4 siloxane was not detected in the urine, rather D4 metabolites accounted for all measured ^{14}C radioactivity. Approximately 95% of the measured ^{14}C radioactivity in the trout bile was attributed to D4 metabolites. The digestive tract and adipose tissue contained high amounts of D4 parent siloxane, while 40% of the radioactivity in the liver was attributed to D4 siloxane metabolites.

D5

- Juvenile rainbow trout ($1.36 \pm 0.29 \text{ g}$ wet weight) were fed ^{14}C -D5 siloxane for 35 d and then allowed to depurate for 42 d (Woodburn et al. 2013, Drottar 2007). The dietary route of exposure represents the most likely exposure route for fish to D4. Whole body measurements from the last three sampling points of the uptake period revealed that the fish achieved steady-state as the residue concentrations did not significantly differ over time. An empirical BMF value for D5 was 0.32. Whole body autoradiography (WBA) was conducted on Days 1 and 10 of the uptake period and Days 2, 14 and 42 of depuration (Drottar et al. 2006 a and b). The concentration of ^{14}C radioactivity in the liver and digestive tract was high throughout the entire study, which is suggestive of metabolism and elimination. After 42 d of depuration, most of the radioactivity was found in the liver and digestive tract (Drottar et al. 2006 a and b).
- Rainbow Trout (500-1000g) were fed ^{14}C - D5 siloxane for five days, which had been mixed into fish feed (Durham et al. 2009) Individual trout were homogenized, extracted with tetrahydrofuran (THF), and then fractionated and analyzed by high performance liquid chromatography (HPLC) to profile potential metabolites. Based on the HPLC metabolite profiling, 31% of the radioactivity in the fish homogenates were D5 siloxane metabolites.
- Rainbow Trout (704 – 1397g) were administered a single orally gavage dose of 15 mg/kg ^{14}C -D5 siloxane (Springer 2007; Domoradzki et al. 2007). Over 96 hr, trout plasma and urine were sampled to determine the amount of parent D5 siloxane and metabolites in each matrix. At the 96 hr terminal sacrifice, trout bile, digestive tract, liver, and fat were also sampled for parent D5 siloxane and metabolites. HPLC metabolite profiling demonstrated that biotransformation products were present in the plasma, urine, bile and digestive tract. Parent D5 siloxane was not detected in the urine, rather D5 metabolites accounted for all measured ^{14}C radioactivity. D5 siloxane metabolites were also present in the bile, in comparison to a small amount of D5 siloxane parent material (99% of radioactivity associated with metabolites).
- In vitro* S9 and microsomal biotransformation of ^{14}C -D5 was studied in a trout, carp and catfish (Cantu et al. 2015). Biotransformation was observed in fish, though was typically $< 5\%$ of the starting concentration.

Expert Opinion

D4

- D4 siloxane is biotransformed by fish. The HPLC metabolite profiling in two *in vivo* studies clearly shows the presence of multiple biotransformation products (Springer 2007; Durham et al. 2009; Domoradzki et al. 2007 and 2009). All of these biotransformation products are more polar than D4 parent siloxane. The presence of these polar metabolites indicates that these metabolites will likely be eliminated quickly from the fish. Further chromatographic data can be found at Domoradzki et al. 2015 (Poster WE224).
- The metabolic rate constant (k_m) for D4 siloxane is $\geq 0.01 \text{ day}^{-1}$. A k_m of 0.14 day^{-1} has been suggested for D4 siloxane and is based on the blood time course data from the D4 rainbow trout gavage study (Domoradzki et al. 2009 a and b). This analysis assumes that the partitioning of D4 between the blood and whole body is identical. While this assumption may not be incorrect based on review of the blood:tissue partitioning data from that study, there is uncertainty around deriving a k_m based on blood time course data versus whole body data. Applying an uncertainty factor of 10 brings the experimentally-derived D4 k_m value to $\geq 0.01 \text{ day}^{-1}$. A value of this magnitude would be consistent with a majority of TMF aquatic food web data on D4, showing biodilution (i.e., TMF < 1).
- The empirical BMF (wet weight basis) for D4 siloxane is < 1 , with hepatic biotransformation playing a minor role. Woodburn et al. (2103) reported an empirical BMF for D4 siloxane of 0.28. In addition, based on the μg dosed and μg recovered in the carcass and tissues from adult rainbow trout at 96 hr in the oral gavage study, a comparable empirical D4 BMF of 0.63 may be calculated (Springer 2007; Domoradzki et al. 2008). We know from multiple studies that D4 siloxane biotransformation products were present in fish after a single oral dose (Durham et al. 2009; Domoradzki et al. 2008 and 2009).
- During D4 siloxane dosing and depuration periods, a majority of the D4 residues are located in the liver, gall bladder and GI tract. The residues being located in the liver and GI tract indicates that the residues are in the process of being cleared from the body.

D5

- D5 siloxane is biotransformed by fish. The HPLC metabolite profiling utilized in the two *in vivo* studies clearly show the presence of multiple biotransformation products (Durham et al. 2009; Springer 2007; Domoradzki et al. 2007). All of these biotransformation products are more polar than D5 parent siloxane. The presence of these polar metabolites indicates that these metabolites will likely be eliminated quickly from the fish.
- The metabolic rate constant (k_m) for D5 siloxane is $\geq 0.01 \text{ day}^{-1}$. In the oral gavage study, approximately 14% of the original D5 siloxane dose was converted to metabolites. Given the trout were dosed orally, the blood was sampled over 96 hr (time points of 0, 2, 4, 8, 12, 24, 48, 72, 96 hr), and both parent and metabolites were measured, a metabolic rate constant (k_m) was estimated from the blood temporal data to be 0.17 day^{-1} (Woodburn and Domoradzki 2008; Domoradzki et al. 2009). This analysis assumes that the partitioning of D5 between the blood and whole body is identical. While this assumption may not be incorrect based on review of the blood:tissue partitioning data from that study, there is uncertainty around deriving a k_m based on blood time course data as opposed to whole body data. Applying an uncertainty factor of 10, brings the experimentally-derived D5 k_m value to $\geq 0.017 \text{ day}^{-1}$. A value of this magnitude would be consistent with a majority of TMF aquatic food web data on D5, showing biodilution (i.e., TMF < 1).
- The empirical BMF for D5 siloxane is < 1 , with hepatic biotransformation seemingly playing a pivotal role. Woodburn et al. (2103) reported an empirical BMF for D5 siloxane of 0.32. In addition, based on the μg dosed and μg recovered in the carcass and tissues of the trout at 96hr in Springer (2007) and Domoradzki et al. (2007), an average empirical BMF of 0.19 may be calculated. We know from multiple studies that D5 siloxane biotransformation products are present in fish after only 4-5 days of oral exposure (Durham et al. 2009; Springer 2007; Domoradzki et al. 2007 and 2009).
- During D5 siloxane dosing and depuration periods, a majority of the D5 residues are located in the liver and GI tract. The residues being located in the liver and GI tract indicates that the residues are in the process of being cleared from the body.
- In vitro* biotransformation data in rainbow trout demonstrate that D5 is converted to more polar products (Cantu et al. 2015).

Conclusion: Based on the available data, there is unequivocal evidence that D4 & D5 siloxane are biotransformed to more polar metabolites in fish. The biotransformation of D4 and D5 siloxanes provide one rationale for biodilution behavior in most natural aquatic food webs (i.e. a TMF < 1). Redman et al. (2012) reported a dietary uptake efficiency of $\sim 10\%$ for D4 and D5 based on measured fish data from Lake Pepin. Goss et al. (2013) suggested that at a 10% dietary uptake efficiency, a threshold for $K_{elimination}$ of 0.001 day^{-1} was needed for a BMF/TMF < 1 . Empirical data support that both D4 and D5 $K_{elimination}$ values are $> 0.01 \text{ day}^{-1}$, which provides underlying rationale for the BMF/TMF for both D4 and D5 being < 1 .